This article was downloaded by: On: 17 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37- 41 Mortimer Street, London W1T 3JH, UK

To cite this Article Leśniewicz, Anna and Żyrnicki, Wiesław(2000) 'Multielemental Analysis of Yew, Pine and Spruce Needles by ICP-AES', International Journal of Environmental Analytical Chemistry, 77: 2, 147 — 158

To link to this Article: DOI: 10.1080/03067310008032678 URL: <http://dx.doi.org/10.1080/03067310008032678>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use:<http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Intern. 1. Enrim. Ad. &a. **Vol. 77(2). pp. 147-158 Rcprinls available dirstly from tk publisher Photocopying permitted by license only**

MULTIELEMENTAL ANALYSIS OF YEW, PINE AND SPRUCE NEEDLES BY ICP-AES

ANNA LESNIEWICZ and **WIESLAW** ZYRNICKI*

Wrockw University of Technology, Chemistry Department, Institute of Inorganic Chemistry and Metallurgy of Rare Elements, Wybrze ie Wyspiańskiego 27, 50-370 Wmchw, Poland

(Received 19 October 1999; lnfinal fom 9 December 1999)

Concentrations of **22** trace and nutritional elements in yew, pine and spruce needles collected from urban and industrial **areas** were determined by inductively coupled plasma atomic emission spectrometry. Dependence of concentrations of trace elements in conifers species on sampling site **was** investigated. **Various** procedures of sample digestion (different acids, conventional hot plate and microwave digestion methods, *dry* ashing) were examined and discussed.

Keywords: Trace and nutritional elements; multielemental analysis; **ICP-AES;** yew; pine; spruce needles

INTRODUCTION

Heavy metals inputs to the biosphere from anthropogenic activities are increasing. Metals enter the biological cycle through the mots and leaves of plants and are enriched in various plant organs. They can directly affect plant growth and an excess dietary intake of contaminated plants is dangerous for the health of humans and animals. The chemical composition of plants is related to the elemental composition of the soil and the contamination of the plant surface indicates the presence of noxious environmental contaminants in ambient **air.** Surface contamination is ascribed to aerosol deposition^[1].

One of the methods of environmental monitoring is assessment of the pollution grounds on chemical composition of plants. For biomonitoring are **used:** mosses, lichen, grass, cultivated plants, deciduous plants and conifers. Different tissues of **trees** such as wood, bark, shoots, buds, cones and needles have been analysed.

^{*} Corresponding author. **Fax: 48-71-328 4330,** E-mail: zymicki@ichn. ch.pwr.wroc.pl

Among conifers Norway spruce *(Picea abies)* ^[2-19] and Scots pine *(Pinus svl* v *estris*) $[4,17,20-23]$ have most often been investigated. Tissues of black pine, loblolly pine, white spruce, red spruce, black spruce, Fraster fir and silver fir [11,24,25, 27-31] have been also studied. Some published papers have been devoted to chemical composition of epicultural waxes and dew from the needles [2,3,5,7,9,32 and ^{33]}. Sampling areas were very different. Samples were collected from open territories $\left[2,3,5,7,8,9,12,16,17\right]$, industrial districts, national parks and mountain forests $[6,11, 22, 24, 29, 31]$ and small areas located around industrial cites, steel mills or generating stations ^[4,20,27,28]. Nutritive elements **(Ca, Mg, K, Na**) and P), heavy metals (mainly: Fe, Cd, **Cu,** Pb and **Zn)** and non-metals **(S,** As, Br and I) have been usually determined. Concentrations of rare-earth elements have also been measured. The INAA ^[2,3,5,7,9,12,24,30,34], AAS ^[6,16,22,24,25,31,35,36,37], XRF ^[8,17], ICP-MS ^[11] and ICP-AES ^{[7,11,17,20,22,28,29,36,38,39] methods have} been employed for determination of elements in needles. Needle samples have been prepared for measurements using **as** a rule wet digestion method. Microwave-assisted method, pressurised decomposition in Teflon bombs and conventional hot-plate digestion in open vessels has been employed. Usually samples are digested in nitric acid $[11,36,37,38]$, nitric acid and hydrogen peroxide $[17]$, nitric acid and perchloric acid $[20,28]$, nitric acid and hydrochloric acid $[31,39]$. nitric acid and sulphuric acid $[31]$ mixtures. Comparison and evaluation of various digestion procedures was performed for some biological and environmental samples, but there is a lack of such a studies for foliage.

The aim of this work was the analysis of trace and nutritional elements in needle samples of yew (not studied so far), pine and spruce from different sites. **Var**ious digestion procedures for needle samples were examined and evaluated. The ICP-AES method was applied for measuring element concentrations.

EXPERIMENTAL

Sampling

Sampling of yew *(Taus boccara),* pine *(Pinus sylvesrris)* and spruce *(Picea ubies* and *Picea galucu)* needles was carried out in November and December in four locations in Lower Silesia (Poland) Wroctaw (a city - **l),** Otawa (an industrial town -2 and 3) and Siechnice (a village 4 -). At each site samples were taken from 3 **to12** trees (depending on number of trees growing in selected location) by cutting branches from the middle of the crown, as described in other studies (see e.g. refs. **7,8, 10,12** and **16).**

Sample pre-treatment

Dust and deposits were removed from the surface of needles by washing in tap and distilled water (see [28]). The cleaned needles were dried in the room temperature (as in Ref. 20,29,40) and removed from twig **axis.** Needles from trees growing at one site were mixed into one laboratory sample (e.g. [12, 401) and stored until analysis in bags.

Reagents

Aqueous standard solutions were prepared by dilution of the stock standard (SPEX Industries) solutions. The following chemicals of analytical grade were used for sample preparation: 65% Nitric Acid - Merck KGaA, Darmstad, Germany; PPH "Standard", Lublin, Poland; **35** - 38% Hydrogen Chloride - PPH "Standard", Lublin, Poland; **⁹⁵**- **97%** Sulphuric Acid - TZPN "Polchem", Toruh, Poland; 30% Hydrogen Peroxide - PPH-PoCh, Gliwice, Poland.

Deionized water (specific resistivity -18.3 M Ω cm) was prepared with the aid of an Easy pure RF (series 703, Barnstead, Thermolyne Corporation, USA) water purification system.

Glassware and plastic bottles were cleaned by soaking them for two days in 10% nitric acid and rising with deionized water.

Sample digestion procedures

Conventional digestion

A portion of 10 g (weighted exactly) of needles was placed in a clean, glassy baker. 100 ml of concentrated nitric acid was added and the baker (covered with a watch glass) was left overnight. The next day, the solution was gently heated on the conventional hot plate. Next portions of nitric acid were added and sample was heated again. The solution was evaporated to a small volume. After cooling hydrogen peroxide was added dropwise and the solution was heated - this step was repeated until the solution was colourless. Finally, the digestion residues were diluted with deionized water to *50* **ml** in the calibrated flask.

Micro *wave-assisted digestion*

About 500mg of powdered spruce needles were accurately weighed into a Teflon digestion vessel. 6 ml of concentrated nitric acid and 1 ml of hydrogen peroxide were added. Decomposition of samples was carried out in a microwave digestion system (Milestone, MLS-1200, MEGA). The following program was

applied to the examined material: step 1- **2** min, **250 W,** step **2- 2** min, **0** W, step 3- 6 min, **250** W; step **4- 5** min, **400** W, step **5- 5** min, **600 W,** step 6- **5** min, ventilation. After cooling the sample solutions were quantitatively transferred into **25** ml volumetric flask and filled up with deionized water to the mark.

Dry ashing

A 10-gram of air-dried needle sample was weighted to a porcelain crucible. Six crucibles were placed in a microwave furnace (MAS **7000,** CEM Corporation) and ashed in three steps with the following times corresponding to the temperature rise and heating at the selected constant temperature: step 1- 100 **"C, 30** min, **5** min; step **2-** 300 **"C, 30** min, **5** min; step **3- 550°C. 45** min, **12** h. The totally ashed sample was moistured with deionized water and dissolved in 6 ml of concentrated nitric acid. Finally, the crucible content was transferred into **50** ml flask.

In the described above digestion procedures, before analysis all the solutions were filtered through the hard filter paper. All investigated sample solutions were prepared in **5%** nitric acid.

Instrumentation

The element concentrations in needle samples were measured using atomic emission spectrometry with inductively coupled argon plasma **as** the excitation source. A Jobin-Yvon **38s** spectrometer working under conditions shown in Table I was used.

For all measurements blank solutions were prepared and taken into account.

RESULTS AND DISCUSSION

Sample homogeneity

The homogeneity of the examined material was studied by analysis of ten replicate samples digested in nitric acid with addition of hydrogen peroxide. The mass of every sample was 10 gram. The results of the study **are** presented in Table **I1** in the form of relative standard deviations for individual elements. As can be observed the **RSD** values for most of the elements are low, indicating that the examined material is quite homogenous.

Discharge parameters:				
	Forward power	1000W		
	Plasma gas flow rate	$13 L min-1$		
	Sheath gas flow rate	$0.20 L min^{-1}$		
	Nebulizer gas flow rate		0.3 L min ⁻¹ (cross-flow nebulizer)	
	Sample uptake	1.5 mL min ⁻¹		
Mnochromator:				
		1 m Czerny-Turner type: HR 1000		
	Gratings	4320 and 2400 grooves mm^{-1}		
	Slit width (entrance/exit)	$20 \mu m / 50 \mu m$		
Analytical lines (wavelengths in nm):				
Al	226.922, 396.152	Mg	279.553, 285.213	
в	249.773	Mn	257.610, 259.373, 294.920	
Ba	233.527, 455.403	Na	588.995	
Ca	317.933, 393.366	Ni	221,647, 231, 604, 232, 003	
Cd	214.438, 226.502, 228.802	P	213.618	
Co	228.616	Pb	220.353	
Сr	205.552, 206.149, 267.716	Sг	407.771	
Cu	224.700, 324.754	Tì.	334.904, 334.941	
Fe	238.204.259.940	v	292.402	
K	766.490	Zn	202.548, 206.200, 213.856, 334.502	

TABLE **1** Instrumental and operating conditions for ICP-AES

~~~~~ ~~~ ~ ~ **TABLE II** Sample homogeneity obtained for ten replicates

| <b>RSD</b> [%] | <b>Elements</b>                  |  |  |
|----------------|----------------------------------|--|--|
| ⊲              | Al, Ca, Fe, B, Cu, Ti, Cd, Cr, V |  |  |
| $2 - 4$        | K, Mg, Ba, Mn, Pb, Zn            |  |  |
| $4 - 6$        | Co, Ni, P, Sr                    |  |  |

# **Evaluation of applicability of various acids for digestion**

The effectiveness of different acid mixtures for the digestion of needle samples was investigated. **To** test the influence of different oxidant mixtures five replicate samples of each material were digested with the following concentrated acid mixtures: nitric acid, nitric and hydrochloric acids, and nitric and sulphuric acids.

The element concentrations measured in solutions obtained after digestion in various acids were compared. For most of the elements their concentrations were found to be consistent **as** shown, for example, for Fe, Mg and P in Figure 1. For a few elements (B, Ba and Pb) different concentrations were obtained (see Figure 1). Meaningful differences have been found for Ba and B. In the case of Ba it can be explained by the formation of hardly soluble  $BaSO<sub>4</sub>$ . The low concentration of B measured in the solutions digested in the  $HNO<sub>3</sub> + H<sub>2</sub>SO<sub>4</sub>$  mixture was rather unexpected. It can be due to various reasons. In addition, for the mix**ture** of nitric and sulphuric acids, the **RSD** values of the concentrations of almost all elements were higher then for other acids. The highest concentrations were measured in the solutions digested with the aid of nitric acid. Therefore, the nitric acid was employed for the next experiments.

### **Comparison of different digestion methods**

Selected results of the determination of trace and mineral elements in grounded spruce needles digested by three different methods are shown in Figure 2. For inorganic matrix elements (Ca, Mg, P) and for most of the other elements (e.g. Ba, B, Cu) the three methods have led to consistent results. However, for Mn, Al, Ti and Cr *dry* ashing has been found to be an ineffective method of needle sample digestion. This can be due to the formation of compounds only partially or hardly soluble, as reported by Hoenig et al. <sup>[41]</sup>.

The microwave-wet decomposition in a high-pressure system **has** been found to be the most efficient method for digestion of such a plant material **as** tree needles. Small amounts of reagents were used. An additional advantage of this method is the very short time of digestion (about **25** min.) if one compares the conventional hot-plate digestion and *dry* ashing methods.

## **Trace and nutritional elements in needles**

Using multielemental standard solutions for calibration, concentrations of Al, B, Ba, Ca, Cd, Co, Cr, Cu, Fe, K, Mg, Mn, Na, Ni, **P,** Pb, Sr, Ti, **V** and Zn were simultaneously measured. During all measurements the argon line *Ar* I **356.766**  nm was monitored to control the stability of plasma excitation conditions.

For Al, Ca, Cd, Cr, Cu, Fe, Mn, Mg, Ni, Ti and Zn in yew, spruce and pine samples, concentrations were measured from a few analytical lines. Results of the element concentration measurements from different emission lines are given



**FIGURE 1 Comparison of needle digestion in various acid mixtures (1-**  $HNO_3$ **, 2-**  $HNO_3$  **+**  $HCl$ **, 3-** $HNO<sub>3</sub> + H<sub>2</sub>SO<sub>4</sub>$ 

Mg

P

200 0

Fe

in Figure **3.** For most of the elements measured consistent concentration values were obtained. Only for **Al** and Cr significant differences in the concentrations were observed. As it has been reported for Al<sup>[42,43]</sup> it can be explained by spectral interferences of the A1 **396.2 nm** line. The selection criteria of lines for final measurements were signal to background ratio, detection limit and spectral interferences.

The final measurements of aluminium and chromium were based on **A1226.9**  nm and Cr **205.5** nm lines.

The matrix effects in the analysis of yew and spruce needles by the **ICP-AES**  method were evaluated by comparison of calibration curve and standard addition



FIGURE 2 Concentrations of elements  $\left[ \text{in } \mu \text{g/g} \right]$  – comparison of different digestion procedures:  $1 - \text{microwave}$ -assisted,  $2 - \text{conventional hot-plate and } 3 - \text{dry ashing}$ 

methods. The standard addition method was performed for the detemination of Al, B, Ba, Cd, Cr, **Cu,** Fe, Ni, Pb, Sr, Ti, **V** and **Zn.** It was found that at appropn**ate** dilutions of the measured solutions the matrix effects are lower than the standard deviation uncertainties of the concentrations.

Analysis of yew needles collected from three different sites **was** carried out. The results obtained (Table III) demonstrate dependence of the elemental composition of needles on the trees **growing** site. The very **high** levels of zinc in needle samples from Otawa (Stations **2** and 3) were connected with the presence of the mill in the town. Generally, in samples **from** urban areas (both Wroctaw and Otawa), the concentrations of Al, B, Ba, Cr, **Cu,** Fe, **Mg,** Pb, Ti and **V** were



**FIGURE 3 Concentration of selected elements measured from various lines in pg/g** 

higher than those found in needles from the place located outside the town, while the lowest concentration of Mn was found in town samples (Stations 1 and **2).** In the samples **from** the centre of Otawa (2) the highest concentrations of Al, Fe, Mg, Pb, and Ti **are** observed. The concentration levels of the trace (B, Ba, Cr, Cu, **Sr** and **V)** and nutritional (Ca, K, **Na,** P) elements are the highest in the samples from Wrocław.



Literature date 1498-6400 0.081-0.753 1800-8973 *Measured Literature data Literature data Literature data* 0.081-0.753 1498-6400 0.074-0.36 800-8973 751-2000 11.3-33.6 0.14547.1 1.2-207.3 0.1-395.6 530-2200 0.12-10.8 25-2555 194-1001 0.13-72 20-43 1 46-500 0.5-0.9 13-480 1.5-16 2-5 *NORWAY SPRUCE (3)" PINE (4)"*   $PNE(4)^{a}$ Measured 7210 Al **126 133 41.8 133 41.8** 133 41.8 133 41.8 141 23.6 2.15 *3ooo*  1450 36.9 <u>105</u> 0.25 0.23 3.73 98.7 465 765 191 2.11 3.23 3.84 1.89 0.17 - *YEW(1)" YEw(2)" YEw(3)" WHITE SPRUCE (3)"*  WHITE SPRUCE (3)<sup>a</sup> 7610 1720 **TABLE** III **Concentrations of elements in needles in pg/g**  18.8 3200 48.5 17.6 47.8 0.53 0.42 4.60 387 533 3.38 21.5 15.2 0.88 63.3 141  $\frac{1}{4}$ 521 Literature data 0.064-0.19 0.1-0.336 0.3 124 **.O**  3400-m 1500-1600 1.0-53.2 6w-ao78 1.75-5.09 16.3-150 220-1500 1.2-3.5 NORWAY SPRUCE (3)<sup>a</sup> 33-3474  $6.8 - 319$ 17-63 0.8-40 40-99 6.9-80 - -  $\vec{\triangledown}$ **a Numbers m brackets correspond to the locations described in the text.**  a. Numbers in brackets correspond to the locations described in the text. Measured 23.8 3700 7560 1630  $\frac{1}{87.7}$ 42.5 0.51 0.33 1.22 3.52 235 17.7 494 0.94 11.8 26.2 9.16 0.58 71.7 45 1  $YEW$  $(3)^d$ 5520 6970 1320 93.0 41.8 14.3 12.0 0.33 18.8 4.75 0.36 0.47 0.84 3.68 *550*  376 1.37 6.92 135 190  $YEW(2)^a$ 0.16 7200 25.4 19.9 1.42 13.9 133 5980 **0.24**  5.53 869 65.3 288 1.38 19.6 0.86 77.7 461  $\mathbf{I}$  $\bar{1}$  $VEW$   $(1)^a$ 11800 1810 126 37.6 23.9 7250 0.50 0.43 1.74 9.40 810 59.5 612 1.29 15.4 9.99 0.98 47.2 432 29.2 **Mg**   $\bar{\mathbf{z}}$ **Na**  *co*  zn ₹ **Ba**  *ca cd cr*   $\vec{c}$ **Fe**   $\Sigma$ <u>ድ</u> **a B K P Sr**   $\triangleright$ 

TABLE III Concentrations of elements in needles in µg/g

# ANNA LESNIEWICZ and WIESŁAW ŻYRNICKI

Results of the analyses carried out on needle samples of three species of conifers picked in the same place in Ofawa (3) are **also** included in Table III. The comparison of concentrations of the elements in yew and two spruce samples from one site and yew needle samples collected from three locations indicates different metal uptake by the examined species. The results obtained here **indi**cate that the species have different characteristics for monitoring pollution. Differences in concentrations for B, Cd, Co, Cu, K, Mg and P are smaller than for Al, Ba, Ca, Fe, **Mn,** Ni, Pb, Sr, Ti, **V** and **Zn.** The highest quantities of trace *(Al,*  Ba, Cd, Co, Cr, Cu, Fe, Ni, Pb, Ti and **V)** and main (K, Na and P) elements **in**  needles of white spruce were measured. The amounts of Ca, Mg, and **Zn** in yew needles are considerably higher than in spruces. The results obtained for spruce and pine needles are comparable with literature data. Only the concentrations of cadmium, chromium, iron, phosphorus and lead in the Norway spruce needles *are* greater.

#### *References*

- [I] A. Kabata-Pendias **and** H. Pendias, "bee *elements in plants and* soils", CRC **Press,** Bota Raton, **Florida (1984);.**
- **[2]** A. Wyttenbach, **S.** Bajo **and** L. Tobler, *Fresenius* J. *Anal. Chem.,* **345,294-297 (1993);.**
- **[31 L.** Tobler. **S.** Bajo, **A.** Wyttenbach **and** M. Langenauer, Fresenius J. *Anal. Chm.,* **350,** *85-88*  **(1994);.**
- **[4]** M. Rossbach **and** R. Jayasekera, Fresenius *J. Anal. Chem.,* **354,511-514 (1996);.**
- **151 A.** Wyttenbach. **S.** Bajo **and** L. Tobler, *Fresenius* J. *Anal. Chem..* **354,668-671 (1996);.**
- $[6]$  M. Kazda and L. Zvacek, *Plant and Soil*, 114, 257-267 (1989);.
- **[7]** A. Wyttenbach, P. Schleppi, L. Tobler, **S.** Bajo **and** J. Bucher, *Phr and Soil,* **168-169,** *305-*  **312 (1995);.**
- **181 J.N. Cape,** P.H. Freer-Smith, LA. Paterson, J.A. Parkinson **and** J. Wolfenden, *lkes,* **41, 211- 224 (1990);.**
- **[9]** A. Wyttenbach, **S.** Bajo, J. Bucher, V. Far, P. Schleppi **and** L. Tobler, Trees, **10, 31-39**  (1 **995);.**
- [ **101** R.F. Hiittl **and** J. Wisniewski, Water; *Air and Soil Pollution.* **33.265-276 (1987);.**
- **Ill]** A. **Santem,** J.M. Mermet **and** V.R. Villanueva, Water; *Air* **and** *Soil Pollution, 52,* **157-174 (1990);.**
- [12] D.R. Sauerbeck, *Water, Air and Soil Pollution, 57-58, 227-237 (1991);.*
- **<sup>1131</sup>**J. Ke **and** J.M. **Skelly,** *Wares Air andsoil Pollution,* **74,289-305 (1994);.**
- **[14] M. Mandere and L.** Tuulmets, Water; *Air andSoil Pollution,* **94,247-258 (1997);.**
- [ **151 A.** Wyttenbach, **S.** Bajo, V. Furrer, M. Langenauer **and** L. Tobler, Water; *Air* **and** *Soil Pollution,*  **94,417430 (1997);.**
- [16] B. Maňkovská, *Lesnictví-Forestry*, **43 (3)**, 117-124 (1997);.
- **1171** J. Boman, C. Larsson, M. **Olsson and** H. Raitio, *X-Ray* Spectrometry, **25,89-94 (19%);.**
- **1181 J. Opydo,** Chem Anal. (Warsaw), **42, 123-1928 (1997);.**
- [19] L. Harju, J.-O. Lill, K.-E. Saarela, S.-J. Heselius, F.J. Hernberg and A. Lindroos, *Fresenius J. Anal. Chem.,* **358,523-528 (1997);.**
- $[20]$  B. Berg, G. Ekbohm, B. Söderström and H. Staaf, Water, Air and Soil Pollution, 59, 165-177 **(1991);.**
- **1211 G. Huhn,** H. Schulc. H.-J. **Stiirk,** R. Tolle **and** G. Schiiiirmann, *Ware6 Air and Soil Pollution,*  **84,367-383 (1995);.**
- **[22]** H.B. Lukina, Lesobodenie. **1-2,41-53 (1996);.**
- **1231 E.** Wieteska, **D.** Zi6tek **and** A. Dnewihska, *Anal. Chim Acta,* **330.251-257 (1996);.**
- **[24]** W.P. Robrage. J.M. Pye **and** R.I. Bruck, *Plant and* Soil, **114, 19-34 (1989);.**
- [25] P.A. Arp, C. Akerley and K. Mellerowicz, Water, Air and Soil Pollution, 48, 277–297 (1989);.<br>[26] J.G. Croch, R.C. Severson and L.P. Gough, Water, Air and Soil Pollution, 63, 253–271 (1992);.
- 
- [27] M. **Dion, S.** Loranger, G. Kennedy. **F.** Courchesne **and** J. Layed, *Wer; Air and Soil Pollution,*  71,2941 (1993);.
- [28] C.L. Carlson **and** C.A. **Carlson.** *Water; Air* and *Soil Pollution,* 72.89-109 (1994);.
- [29] L.P. **Gough.** R.C. Severson **and** L.L. Jackson, *Water; Air &Soil Pollution,* 74.1-17 (1994);.
- [30] Z.Q. **Lin,** P.H. Schuepp, RS. Schemenauer **and G.G.** Kennedy, *Water; Air* **and** *Soil Pollution,*  81, 175-191 (1995);.
- [31] M.R. Shepard, C.E. Lee, R.S. Woosley and D.J. Butcher, *Microchemical Journal*, 52, 118-126 9 1995);.
- [32] J. Hovorka and G.B. Marshall, *Fresenius J. Anal. Chem.*, 358, 635-640 (1997);.
- [33] J. Burkharadt and R. Eiden, *Trees*, 4, 22-26 (1990);.
- [34] M.C. Freitas, *Fresenius J. Anal. Chem.*, 352, 58-60 (1995);.
- [35] D. **Chakraborti,** M. Burguera **and** J.L. Burguera, *Fmsenius J. Anal. Chem.,* 347, 233-237 (1993);.
- [36] D.C. Baxter, R Nichol, D. Littlejohn, Ch. Liidke, J. Skole **and** E. **Hoffmaan,** JAAS, 7,727-734 (1992);.
- [37] C.M.M. Smith, N. Robbin **and D.** Littlejohn, JAAS, 8,989-994 (1993);.
- [38] C. Mohl **and** M. Stoeppler, *Fresenius J. Anal.* Chem., 345,164-165 (1993);.
- [39] S.R. Koirtyohann **and D.A. Yates,** *Atomic Spectroscopy,* 15,167-168 (1994);.
- [40] E.M. Lobersli **and** E. Steines, *Water; Air and Soil Pollution,* 37.25-29 (1988);.
- [411 M. Hoenig, H. **Baeten, S.** Vanhentemijk, E. Vassileva **and** Ph. Quevauviller, *Anal. Chim Acta,*  358.85-94 (1998);.
- [42] T.J. Burden, J.J. Powell **and** RP. Thompson, JAAS, 10,259-266 (1995);.
- 1431 S. Reclmagel, U. RBsich **and** P. Briltter, JAAS, 9,1293-1297 (1994).